

# Elevated CO<sub>2</sub> increases shoot growth but not root growth and C:N:P stoichiometry of *Suaeda aralocaspica* plants

WANG Lei<sup>1,2\*</sup>, FAN Lianlian<sup>1,3</sup>, JIANG Li<sup>1,2</sup>, TIAN Changyan<sup>1,2</sup>

<sup>1</sup> State Key Laboratory of Desert and Oasis Ecology, Xinjiang Institute of Ecology and Geography, Chinese Academy of Sciences, Urumqi 830011, China;

<sup>2</sup> University of Chinese Academy of Sciences, Beijing 100049, China;

<sup>3</sup> Research Center for Ecology and Environment of Central Asia, Chinese Academy of Sciences, Urumqi 830011, China

**Abstract:** The purpose of the current study was to investigate the eco-physiological responses, in terms of growth and C:N:P stoichiometry of plants cultured from dimorphic seeds of a single-cell C<sub>4</sub> annual *Suaeda aralocaspica* (Bunge) Freitag and Schütze under elevated CO<sub>2</sub>. A climatic chamber experiment was conducted to examine the effects of ambient (720 µg/L) and CO<sub>2</sub>-enriched (1440 µg/L) treatments on these responses in *S. aralocaspica* at vegetative and reproductive stages in 2012. Result showed that elevated CO<sub>2</sub> significantly increased shoot dry weight, but decreased N:P ratio at both growth stages. Plants grown from dimorphic seeds did not exhibit significant differences in growth and C:N:P stoichiometric characteristics. The transition from vegetation to reproductive stage significantly increased shoot:root ratio, N and P contents, but decreased C:N, C:P and N:P ratios, and did not affect shoot dry weight. Moreover, our results indicate that the changes in N:P and C:N ratios between ambient and elevated CO<sub>2</sub> are mainly caused by the decrease of N content under elevated CO<sub>2</sub>. These results provide an insight into nutritional metabolism of single-cell C<sub>4</sub> plants under climate change.

**Keywords:** biomass; CO<sub>2</sub> elevation; C:N:P stoichiometry; seed heteromorphism; *Suaeda aralocaspica*

## 1 Introduction

Since First Industrial Revolution, atmospheric CO<sub>2</sub> has steadily increased, and current concentration is above 720 µg/L and could reach 1440 µg/L by the year 2100 (IPCC, 2013; Betts et al., 2016). Such elevation in atmospheric CO<sub>2</sub> induces several alterations in plant metabolism. Growth and ecological responses of terrestrial plants to elevated CO<sub>2</sub> have been determined on three main types of photosynthesis: C<sub>3</sub>, C<sub>4</sub> and crassulacean acid metabolism (CAM) types (Ceusters and Borland, 2011; Reich et al., 2018; Quirk et al., 2019). Responses of different plant functional types differ to elevated CO<sub>2</sub>. In many C<sub>3</sub> and CAM plants, plant growth is stimulated by elevated CO<sub>2</sub> (Ceusters and Borland, 2011; Boretti and Florentine, 2019; Li et al., 2020). However, C<sub>4</sub> plants show little or no increase in productivity with elevated CO<sub>2</sub> (Boretti and

\*Corresponding author: WANG Lei (E-mail: egiwang@ms.xjbg.ac.cn)

The first author and the second author contribute equally to this work.

Received 2021-08-23; revised 2021-11-19; accepted 2021-11-24

© Xinjiang Institute of Ecology and Geography, Chinese Academy of Sciences, Science Press and Springer-Verlag GmbH Germany, part of Springer Nature 2021

Florentine, 2019). So far, there is no information available regarding responses of  $C_4$  species without Kranz anatomy to atmospheric  $CO_2$  elevation.

Some plants can produce heteromorphic seeds that are distinctly different in morphology, color, dormancy and germination traits (Imbert, 2002; Zhao et al., 2018). Plants cultured from heteromorphic seeds can also differ in their growth responses under different growth conditions, including biotic and abiotic factors (Redondo-Gómez et al., 2008; Jiang et al., 2020). However, the effects of elevated  $CO_2$  on growth, ecological and physiochemical characteristics are still unknown for seed-heteromorphic plants.

Carbon (C), nitrogen (N) and phosphorus (P) contents and C:N:P ratio is associated with plant adaptation strategy to environmental changes (Du et al., 2019; Li et al., 2021). C, N and P provide the structural basis of plants and play essential role in photosynthesis and growth (Marschner, 2013). Though these elements are maintained at relative cellular concentration in plants, C:N:P stoichiometry of plants can be greatly affected under different environmental conditions (Deng et al., 2015). Numerous simulation experiments were carried out to study the response of elevated  $CO_2$  on plant C:N:P stoichiometry (Pérez-Romero et al., 2018; Du et al., 2019; Uchytlová et al., 2019). However, the effects of elevated  $CO_2$  remain inconsistent, for example, an average of 22% increase in the foliar C:N ratio in  $C_3$  grasses and forbs was observed under high  $CO_2$  concentration, but no significant effects were observed in  $C_4$  grasses (Sardans et al., 2012).

*Suaeda aralocaspica* (Bunge) Freitag and Schütze is a habitant of the saline and semi-arid deserts in Central Asia (Commissione Redactorum Florae Xinjiangensis, 1994). This plant does not have dual-cell (Kranz) system, but has unusual mechanisms of  $C_4$  photosynthesis by dimorphic chloroplasts in individual chlorenchyma cells via intracellular compartmentation (Voznesenskaya et al., 2001). In *S. aralocaspica*, photosynthetic chloroplasts are assembled at the proximal end in the flower and leaves, while other chloroplasts positioned and partitioned on the fringe of the cells at their distal end. In  $C_4$  cycles, atmospheric  $CO_2$  is fixed via phosphoenolpyruvate carboxylase (PEPC) at the distal end of the cell, which produces  $C_4$  acids and donates  $CO_2$  to rubisco in chloroplasts at the proximal end of the cell (Boyd et al., 2007).

We conducted a chamber experiment to examine the growth and C:N:P stoichiometry for plants grown from dimorphic seeds of this single-cell  $C_4$  species to elevated  $CO_2$ . We hypothesized that plants from dimorphic seeds might have differential eco-physiological responses to  $CO_2$  elevation at different growth stages.

## 2 Materials and methods

### 2.1 Seed collection and experimental design

Freshly matured *S. aralocaspica* fruits were harvested from the southern edge of the Junggar Basin in Xinjiang Uygur Autonomous Region, China, in October 2011. Fruits were naturally dried for 2 weeks at ambient room conditions. Dry fruits were rubbed by hand and both brown and black seeds were separated and stored in plastic bags at room temperature (20°C–25°C).

A pot experiment was conducted in two growth chambers (day/night air temperatures: 25°C/10°C for the first three months and then 30°C/15°C) at Fukang National Field Scientific Observation and Research Station for Desert Ecosystems, Chinese Academy of Sciences (44°17'N, 87°56'E; 460 m a.s.l.) in Xinjiang, China. The average concentration of  $CO_2$  for growth chambers was either 720 or 1440  $\mu g/L$ . The  $CO_2$  levels were continuously controlled with an automatic  $CO_2$  supply system (ESHENGTAIHE CTRL TECH, YB6, Beijing, China).

Twenty seeds of each brown and black seeds were sown into plastic pots (17 cm deep and 16 cm in diameter) filled with 2 L mix of quartz sand and vermiculite (3:1 v/v) at a soil depth of 2 mm. Two weeks after, one seedling was kept per pot, and seedlings with similar height and growth were kept for further experimentation. In each pot, 6 g of Osmocote 301 (Scotts, Marysville, OH, USA) was applied prior to sowing and 100 mL of Peters 1 nutrient solution was applied once a week. Salt stress was imposed at 4 weeks after sowing using a mixed salt  $NaCl:Na_2SO_4:NaHCO_3$  (20:20:1). Each pot received 100 mL salt solution (1 g/L) once a week

(Wang et al., 2012). Six plants grown from each seed type were harvested for the first time 1157 under each CO<sub>2</sub> level on 15 July 2012. The second harvest took place on 15 September 2012.

## 2.2 Data collection

### 2.2.1 Plant biomass

After harvesting, shoot and root parts were separated and dried in the oven at 80°C for 48 h to reach a constant weight. Shoot and root biomasses were measured by an electronic analytical balance.

### 2.2.2 C, N and P content

Before chemical analysis, the dried shoot materials were ground in a ball mill, and then passed through a 0.25-mm sieve. The ball mill was cleaned completely after each sample milling. The shoot C content was determined using titration method after digested in K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub>-H<sub>2</sub>SO<sub>4</sub> (Bao, 2000). N content was determined using Kjeldahl method after digestion in H<sub>2</sub>SO<sub>4</sub>-HClO<sub>4</sub> (Lin et al., 2011). P content was determined by colorimetric analysis (formation of chloro-phosphoric molybdate) after digestion in H<sub>2</sub>SO<sub>4</sub>-HClO<sub>4</sub> (Ren et al., 2018).

## 2.3 Statistical analyses

All data were expressed as mean±standard error. The data meet the assumptions of normality and homoscedasticity. Split-plot analysis of variance (ANOVA) was used to analyze the effects of CO<sub>2</sub> level and seed type on shoot and root dry weights, shoot C, N and P contents, and their ratios in *S. aralocaspica* plants, with CO<sub>2</sub> level (720 or 1440 µg/L) as main plot and seed type (brown and black seeds) as subplot at vegetative or reproductive stage. All data analyses were performed using SPSS v13.0 (SPSS Inc., Chicago, IL, USA).

## 3 Results

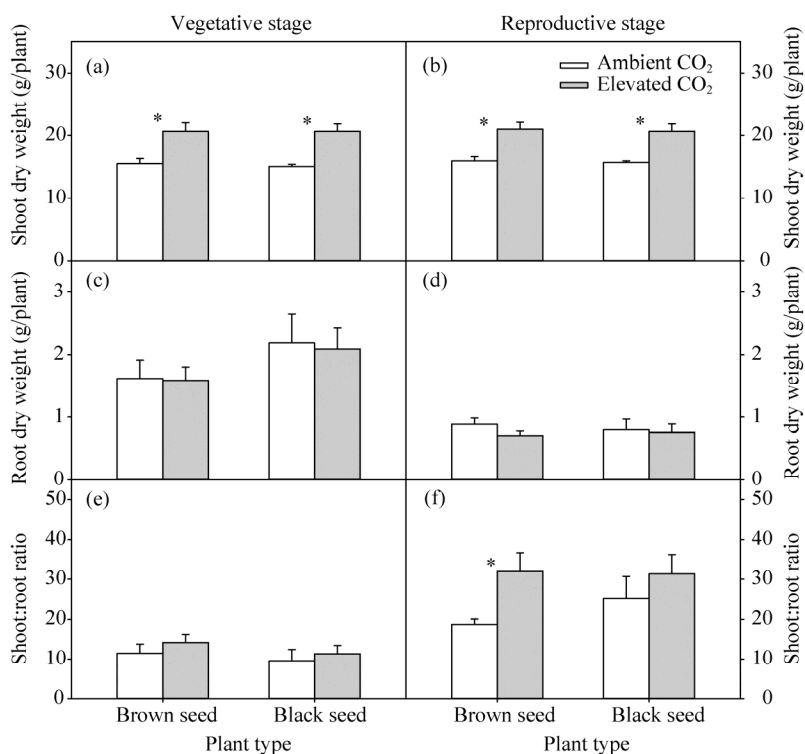
Elevation of CO<sub>2</sub> significantly increased shoot dry weight (SDW) both at vegetative and reproductive stages, but did not affect root dry weight (RDW) and shoot:root ratio. Plants grown from dimorphic seeds showed similar SDW, RDW and shoot:root ratio at the same growth stage under the same CO<sub>2</sub> condition (Fig. 1; Table S1). RDW decreased and shoot:root ratio increased during the transition from vegetative to reproductive stage (Fig. 1).

Elevation of CO<sub>2</sub> significantly decreased N content for plants grown from black seeds at vegetative stage, and did not affect other stoichiometric characteristics. Plants cultured from dimorphic seeds had the same C, N and P contents for both CO<sub>2</sub> treatments at the same growth stage (Fig. 2; Table S1). N and P contents in shoot significantly increased from vegetative stage to reproductive stage, however C content significantly decreased during this transition (Fig. 2). The range of C contents was between 309.4 and 330.4 mg/g. Plants from black seeds at elevated CO<sub>2</sub> showed the highest N (22.8 mg/g) and P content (3.7 mg/g) at reproductive stage (Fig. 2d and f).

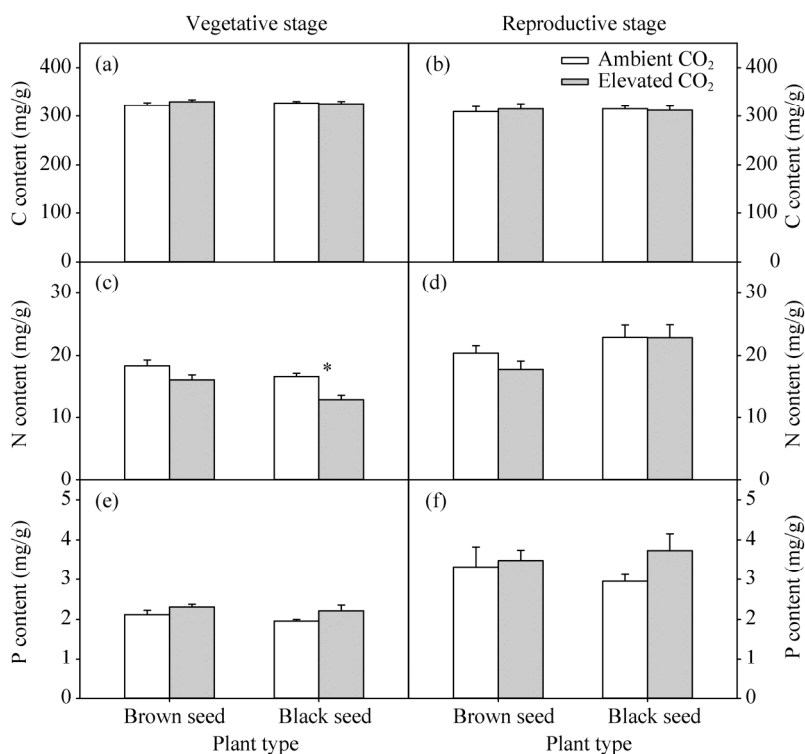
Plants grown from dimorphic seeds had the same C:N, N:P or C:P ratio at same growth stage under the same CO<sub>2</sub> condition (Fig. 3). Compared with vegetative stage, plants at reproductive stage had lower C:N, C:P and N:P ratios (Table S1). Plants grown from black seeds at elevated CO<sub>2</sub> showed the lowest C:N ratio (14.3) and C:P (89.7) at reproductive stage (Fig. 3b and d). N:P ratio significantly decreased under elevated CO<sub>2</sub> in both plant types at vegetation stage (Fig. 3e).

## 4 Discussion

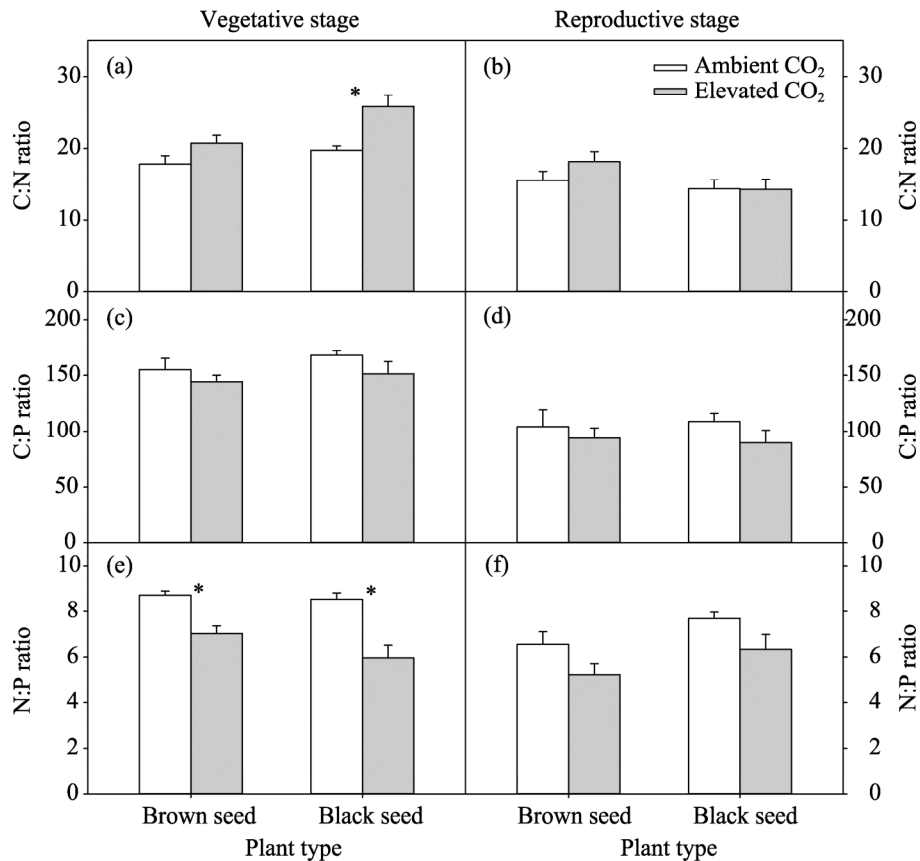
Although the growth and C:N:P stoichiometry of plants with different photosynthetic pathways subjected to elevated CO<sub>2</sub> have been studied extensively (Sardans et al., 2012; Deng et al., 2015; Du et al., 2019), our data are the first in which responses to CO<sub>2</sub> elevation have been documented for single-cell C<sub>4</sub> plants. In addition, our data indicate that elevated CO<sub>2</sub> significantly increases the shoot growth at both vegetative and reproductive stages, but not the root growth and C:N:P stoichiometry. The results also indicate that plants cultured from dimorphic seeds of *S. aralocaspica* have similar growth and C:N:P stoichiometry.



**Fig. 1** Effects of elevated CO<sub>2</sub> on shoot dry weight (a and b), root dry weight (c and d), and shoot:root ratio (e and f) in plants grown from dimorphic seeds of *Suaeda aralocaspica* at vegetative and reproductive stages. \* indicates significant difference between ambient and elevated CO<sub>2</sub> treatments at  $P < 0.05$  level.



**Fig. 2** Effects of elevated CO<sub>2</sub> on C content (a and b), N content (c and d), P content (e and f) in shoots of plants grown from dimorphic seeds of *Suaeda aralocaspica* at vegetative and reproductive stages. \* indicates significant difference between ambient and elevated CO<sub>2</sub> treatments at  $P < 0.05$  level.



**Fig. 3** Effects of elevated CO<sub>2</sub> on C:N ratio (a and b), C:P ratio (c and d), and N:P ratio (e and f) in shoots of plants grown from dimorphic seeds of *Suaeda aralocaspica* at vegetative and reproductive stages. \* indicates significant difference between ambient and elevated CO<sub>2</sub> treatments at  $P < 0.05$  level.

Elevated CO<sub>2</sub> alters plant growth, contents of C, N and P, and C:N and N:P ratios (Li et al., 2019). Our results confirmed that elevated CO<sub>2</sub> significantly accelerated shoot growth and decreased N content of *S. aralocaspica*, which was consistent with previous studies. On average, N content in plant tissue decreases by about 14% under elevated CO<sub>2</sub> (Cotrufo et al., 1998). Elevated CO<sub>2</sub> has different effects on the N contents of different plant types and the reduction in N contents is smaller in C<sub>4</sub> plants than in C<sub>3</sub> plants. The changes in C:N and N:P ratios of *S. aralocaspica* between ambient and elevated CO<sub>2</sub> are mainly caused by the decrease of N content at elevated CO<sub>2</sub>. The results coincide with previous results that C:N ratio increases under elevated CO<sub>2</sub> regimes (Sardans et al., 2012).

Differences exist not only in seed size and germination of heteromorphic seeds but also in plant growth characteristics and even maintain until the end of growing season (Imbert, 2002). Contrary to owning differences in seed dormancy, germination and plant reproduction (Wang et al., 2008, 2012), plants cultured from dimorphic seeds of *S. aralocaspica* did not significantly differ in their responses to CO<sub>2</sub> elevation. The differential response of heteromorphic seeds is effective in reducing the offspring survive risk in unpredictably fluctuating environments. For example, dimorphic seeds of *S. aralocaspica* show different salt tolerance (Wang et al., 2008). Because the steady increase of CO<sub>2</sub> concentrations is the same for plants grown from dimorphic seeds of *S. aralocaspica*, it is not surprising that both plant types show similar growth and C:N:P stoichiometric characteristics.

Our results showed that N and P contents increased with plant developmental stage. This could be explained by that plants at reproductive stage need a higher nutrient concentration to support

fruiting (Marschner, 2013). C content of *S. aralocaspica* was significantly lower than that (46.4% C) of global terrestrial plants (Elser et al., 2000). Decrease of C allocation at reproductive stage could be related to a high salt accumulation of this species.

## 5 Conclusions

Our results reinforce the conclusion that there is no difference between plants grown from heteromorphic seeds in the responsiveness of ecophysiology to steady environmental factors. Elevated CO<sub>2</sub> greatly decreased N content and then caused the change of N:P ratio of *S. aralocaspica*. Our results could enrich the plant response pattern of different types of photosynthesis to CO<sub>2</sub> elevation. If we want to accurately predict responses of various plants under future CO<sub>2</sub> concentration, further work is necessary to explore representative plants of special plant photosynthetic types, such as single-cell C<sub>4</sub> and C<sub>3</sub>–C<sub>4</sub> intermediate plant species.

## Acknowledgements

This research was supported by the Strategic Priority Research Program of Chinese Academy of Sciences (XDA2003010302), the National Natural Science Foundation of China (32171514) and the State Key Laboratory of Desert and Oasis Ecology, Xinjiang Institute of Ecology and Geography, Chinese Academy of Sciences (E1510107).

## References

- Bao S D. 2000. Soil Chemistry and Agriculture Analysis. Beijing: China Agriculture Press, 36–37. (in Chinese)
- Betts R A, Jones C D, Knight J R, et al. 2016. El Nino and a record CO<sub>2</sub> rise. *Nature Climate Change*, 6: 806–810.
- Boretti A, Florentine S. 2019. Atmospheric CO<sub>2</sub> concentration and other limiting factors in the growth of C<sub>3</sub> and C<sub>4</sub> plants. *Plants*, 8(4): 92, doi: 10.3390/plants8040092.
- Boyd C N, Franceschi V R, Chuong S D X, et al. 2007. Flowers of *Bienertia cycloptera* and *Suaeda aralocaspica* (Chenopodiaceae) complete the life cycle performing single-cell C<sub>4</sub> photosynthesis. *Functional Plant Biology*, 34: 268–281.
- Ceusters J, Borland A M. 2011. Impacts of elevated CO<sub>2</sub> on the growth and physiology of plants with crassulacean acid metabolism. In: Lüttge U, Beyschlag W, Büdel B. et al. *Progress in Botany 72*. Berlin Heidelberg: Springer-Verlag, 163–181.
- Commissione Redactorum Florae Xinjiangensis. 1994. *Flora Xinjiangensis*. Urumqi: Xinjiang Science & Technology & Hygiene Publishing House, 57–57. (in Chinese)
- Cotrufo M F, Ineson P, Scott A. 1998. Elevated CO<sub>2</sub> reduces the nitrogen concentration of plant tissues. *Global Change Biology*, 4(1): 43–54.
- Deng Q, Hui D, Luo Y, et al. 2015. Down-regulation of tissue N:P ratios in terrestrial plants by elevated CO<sub>2</sub>. *Ecology*, 96(12): 3354–3362.
- Dijkstra F A, Pendall E, Morgan J A, et al. 2012. Climate change alters stoichiometry of phosphorus and nitrogen in a semiarid grassland. *New Phytologist*, 196(3): 807–815.
- Du C, Wang X, Zhang M, et al. 2019. Effects of elevated CO<sub>2</sub> on plant C-N-P stoichiometry in terrestrial ecosystems: A meta-analysis. *Science of the Total Environment*, 650: 697–708.
- Elser J J, Fagan W F, Denno R F, et al. 2000. Nutritional constraints in terrestrial and freshwater food webs. *Nature*, 408: 578–580.
- Imbert E. 2002. Ecological consequences and ontogeny of seed heteromorphism. *Perspectives in Plant Ecology, Evolution and Systematics*, 5(1): 13–36.
- IPCC. 2013. *Climate Change 2013: The physical science basis*. In: Stocker T F, Plattner G K, Tignor M, et al. *Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge, United Kingdom and New York: Cambridge University Press, 1535.
- Jiang L, Tanveer M, Han W, et al. 2020. High and differential strontium tolerance in germinating dimorphic seeds of *Salicornia europaea*. *Seed Science and Technology*, 48(2): 231–239.
- Li B, Feng Y, Zong Y, et al. 2020. Elevated CO<sub>2</sub>-induced changes in photosynthesis, antioxidant enzymes and signal transduction enzyme of soybean under drought stress. *Plant Physiology and Biochemistry*, 154: 105–114.
- Li T, Su J, Fu Z. 2021. Halophytes differ in their adaptation to soil environment in the Yellow River Delta: Effects of water source, soil depth, and nutrient stoichiometry. *Frontiers in Plant Science*, 12: 675921, doi: 10.3389/fpls.2021.675921.

- Li Y, Yu Z, Yang S, et al. 2019. Impact of elevated CO<sub>2</sub> on C:N:P ratio among soybean cultivars. *Science of the Total Environment*, 694: 133784, doi: 10.1016/j.scitotenv.2019.133784.
- Lin C, Yang Y, Guo J, et al. 2011. Fine root decomposition of evergreen broadleaved and coniferous tree species in midsubtropical China: dynamics of dry mass, nutrient and organic fractions. *Plant and Soil*, 338: 311–327.
- Marschner P, 2013. *Marschner's Mineral Nutrition of Higher Plants* (3<sup>rd</sup> ed). Beijing: Science Press, 135–189.
- Pérez-Romero J A, Idaszkin Y L, Barcia-Piedras J, et al. 2018. Disentangling the effect of atmospheric CO<sub>2</sub> enrichment on the halophyte *Salicornia ramosissima* J. Woods physiological performance under optimal and suboptimal saline conditions. *Plant Physiology and Biochemistry*, 127: 617–629.
- Quirk J, Bellasio C, Johnson D A, et al. 2019. Response of photosynthesis, growth and water relations of a savannah-adapted tree and grass grown across high to low CO<sub>2</sub>. *Annals of Botany*, 124(1): 77–90.
- Redondo-Gómez S, Mateos-naranjo E, Cambrollé J, et al. 2008. Carry-over of differential salt tolerance in plants grown from dimorphic seeds of *Suaeda splendens*. *Annals of Botany*, 102: 103–112.
- Reich P B, Hobbie S E, Lee T D, et al. 2018. Unexpected reversal of C<sub>3</sub> versus C<sub>4</sub> grass response to elevated CO<sub>2</sub> during a 20-year field experiment. *Science*, 360(6383): 317–320.
- Ren C J, Zhang W, Zhong Z K, et al. 2018. Differential responses of soil microbial biomass, diversity, and compositions to altitudinal gradients depend on plant and soil characteristics. *Science of the Total Environment*, 610–611: 750–758.
- Sardans J, Rivas-Ubach A, Peñuelas J, 2012. The C:N:P stoichiometry of organisms and ecosystems in a changing world: A review and perspectives. *Perspectives in Plant Ecology, Evolution and Systematics*, 14(1): 33–47.
- Uchytlová T, Krejza J, Veselá B, et al. 2019. Ultraviolet radiation modulates C:N stoichiometry and biomass allocation in *Fagus sylvatica* saplings cultivated under elevated CO<sub>2</sub> concentration. *Plant Physiology and Biochemistry*, 134: 103–112.
- Voznesenskaya E V, Franceschi V R, Kiirats O, et al. 2001. Kranz anatomy is not essential for terrestrial C<sub>4</sub> plant photosynthesis. *Nature*, 414: 543–546.
- Wang L, Huang Z Y, Baskin C C, et al. 2008. Germination of dimorphic seeds of the desert annual halophyte *Suaeda aralocaspica* (Chenopodiaceae), a C<sub>4</sub> plant without Kranz anatomy. *Annals of Botany*, 102(5): 757–769.
- Wang L, Baskin J M, Baskin C C, et al. 2012. Seed dimorphism, nutrients and salinity differentially affect seed traits of the desert halophyte *Suaeda aralocaspica* via multiple maternal effects. *BMC Plant Biology*, 12: 170, doi: 10.1186/1471-2229-12-170.
- Zhao Y, Yang Y, Song Y, et al. 2018. Analysis of storage compounds and inorganic ions in dimorphic seeds of euhalophyte *Suaeda salsa*. *Plant Physiology and Biochemistry*, 130: 511–516.



## Appendix

**Table S1** Split plot variance analysis of the effects of elevated CO<sub>2</sub>, plant type and their interactions on growth and C:N:P ratio

Growth stage	Index	CO <sub>2</sub>		Plant type		CO <sub>2</sub> ×Plant type	
		<i>F</i>	<i>P</i>	<i>F</i>	<i>P</i>	<i>F</i>	<i>P</i>
Vegetative	Shoot	474.824	0.000*	0.031	0.864	0.034	0.858
	Root	0.046	0.839	4.020	0.073	0.016	0.902
	Shoot:root ratio	1.996	0.217	2.012	0.186	0.075	0.790
	C	0.897	0.387	0.003	0.956	0.947	0.353
	N	6.187	0.055	1.553	0.241	0.082	0.781
	P	4.076	0.099	1.679	0.224	0.002	0.963
	C:N ratio	6.031	0.058	2.021	0.186	0.132	0.724
	C:P ratio	3.937	0.104	1.398	0.264	0.111	0.746
	N:P ratio	28.070	0.003*	0.714	0.418	0.108	0.749
Reproductive	Shoot	51.556	0.001*	0.024	0.880	0.367	0.558
	Root	0.660	0.454	0.069	0.798	0.001	0.972
	Shoot:root ratio	4.184	0.096	0.483	0.503	0.737	0.411
	C	0.034	0.861	0.022	0.884	0.163	0.695
	N	1.652	0.255	0.026	0.876	0.791	0.395
	P	0.565	0.486	3.404	0.095	0.397	0.543
	C:N ratio	0.967	0.371	2.448	0.149	0.698	0.423
	C:P ratio	1.434	0.285	0.001	0.982	0.147	0.710
	N:P ratio	16.494	0.010*	4.097	0.070	0.001	0.971

Note: C, carbon; N, nitrogen; P, phosphorous. \* indicates the significant difference at  $P < 0.05$  level.